Journal of Sports Physiology and Athletic Conditioning (J.S.P.A.C) 2024; 4 (14): 1-11

Research Article

Aerobic exercise affects mitochondrial quality control

Frank Amini¹, Mohammad Ali Azarbayjani ^{2*}, Shahin Riyahi Malayeri ³, Lida Moradi⁴

- 1. Department of Physical Education and Sport Sciences, North Tehran Branch, Islamic Azad University, Tehran, Iran.
- 2. Department of Exercise Physiology, Central Tehran Branch, Islamic Azad University, Tehran, Iran.
- 3. Department of Physical Education and Sports Sciences, East Tehran Branch, Islamic Azad University, Tehran, Iran.
- 4. Department of Physical Education and Sports Sciences, North Tehran Branch, Islamic Azad University, Tehran, Iran.

Received: 12 September 2024 Revised: 25 August 2024 Accepted: 14 November 2024

Keywords:

Mitochondrial unfolded protein response, mitochondrial dynamics (fission and fusion), mitophagy, aerobic exercise

Abstract

Mitochondria play a significant role in morphological and physiological aspects of cells according to their known biological role. Any disruption in mitochondrial function weakens cell function and causes many cell diseases. On the other hand, mitochondria health is associated with cellular efficiency and causes health and physical performance. Evidence shows that one of the molecular mechanisms that regular physical activities, especially aerobic exercises, lead to health is mitochondrial health. Aerobic exercise can regulate and control the mitochondrial unfolded protein response (UPRmt), mitochondrial dynamics (fission and fusion) and mitochondrial. It seems that regulation of mRNA expression of proteins responsible for MQC signaling pathways, reduction of inflammation and oxidative stress is one of the mechanisms by which aerobic exercise benefits mitochondria.

*Corresponding author: Mohammad Ali Azarbayjani
 Address: Department of Exercise Physiology, Central Tehran Branch, Islamic Azad University, Tehran, Iran.
 Email: <u>m azarbayjani@iauctb.ac.ir</u> Tell: +982122481622
 M A A: 0000-0002-3502-7487



1. Introduction

Regular physical activities, especially aerobic exercises, have health benefits. Evidence shows that aerobic exercise is one of the most effective non-pharmacological interventions to enhance health and is an integral part of human life, which has significant effects on maintaining health and preventing health disorders (1,2). A combination of aerobic exercise changes the cellular function, improves the functioning of the organs, and ultimately improves health. Reducing inflammation (3), oxidative stress (4), pathological apoptosis (5), endoplasmic reticulum stress (6), and improvement of mitochondrial function (7) are some of the cellular mechanisms through which aerobic exercise maintains and enhances health. In recent years, researchers have focused on mitochondria's role in improving cell functions. Mitochondria are intracellular organelles that evolved from bacteria endosymbiosis with eukaryotic characteristics (8). Mitochondria are unique, highly dynamic organelles that extract energy from energy sources through aerobic metabolism. They also control fatty acid synthesis, the production of intracellular reactive oxygen species (ROS), oxidative phosphorylation, thermogenesis, and intracellular calcium homeostasis. However, it the regulatory center of cell signal is transmission and controls many intracellular processes (9). Mitochondria's cellular roles are so diverse that its energy-generating role, which has made it the powerhouse of the cell, is one of the countless biological roles of this remarkable organelle (10). Due to mitochondria's multiple roles, any dysfunction in its function results in dysfunction in cells. This is one of the key mechanisms of various diseases and pathologies characterized by dysfunctional mitochondria (11).

Mitochondrial dysfunction is significantly related to a wide range of diseases including diseases, neurodegenerative metabolic disorders, cancer and many other diseases (12). Mitochondria's functional role in health and diseases has drawn researchers to this organelle more than other parts of the cell. The results of the studies of the last two decades show that aerobic exercises can improve health bv affecting the structure and function of mitochondria (13). The evidence shows that aerobic exercises can lead to the maintenance and development of mitochondrial function and the development of health through the effect on the mitochondrial quality control system (MQC) (14,15). The three main mitochondrial quality control (MQC) mechanisms that regulate mitochondrial integrity and maintain mitochondrial function include the activation of mitochondrial unfolded protein response (UPRmt), mitochondrial dynamics (fission and fusion) and mitochondrial mitophagy (16). In this article, the effect of aerobic exercise on mitochondrial quality control mechanisms is investigated.

The effect of aerobic exercise on mitochondrial unfolded protein response

Mitochondrial unfolded protein response (UPRmt) is a critical pathway that maintains mitochondrial homeostasis and proteostasis under stress by stimulating the transcription of nuclear-encoded genes that protect and support mitochondria, and Facilitates cellular adaptation in response to pervasive mitochondrial stress. UPRmt mitochondrial activates specific chaperones and proteases, which results in protein folding or removal of mitochondrial proteins damaged by mtROS. This results in the preservation of mitochondrial quality control (17,18). Mitochondrial chaperone activity is critical for proper folding of folded and unfolded proteins in mitochondria. While proteases break down folded proteins to maintain mitochondrial proteostasis (19). Aerobic training has been studied in different tissues, especially skeletal muscle. One of the most significant features of mitochondria is the high flexibility of this organelle in response to aerobic exercise. This flexibility includes adaptations to their volume, structure and function, which leads to MOC (20). The evidence obtained from the studies shows that aerobic exercise has a significant effect on the activation of the UPRmt pathway, especially in skeletal muscle. The results of the studies show that aerobic exercise activates this pathway by affecting mRNA and proteins related to UPRmt. Exercise on a treadmill increased transcription factor Jun (c-Jun), heat shock protein 60 (HSP60) and CLpP (CLpP) in male C57BL/6J mice, which indicates activation of the classical UPRmt pathway (21). UPRmt pathway proteins decrease with age. Reported in rodents, mRNA levels of YME1L1 and CLpP of gastrocnemius muscle increased significantly

along with mitochondrial content after aerobic training (22). These findings were confirmed in another study which showed that aerobic training enhanced skeletal muscle Yme1L1 and LONP1 mRNA levels along with PGC-1a and citrate synthase levels (23). These findings show that aerobic exercise can enhance skeletal muscle mitochondrial function in old age by stimulating the expression of UPRmt pathway proteins. It can also improve skeletal muscle function, which can increase rip strength, maximum running speed, and running distance. In obese mice fed high-fat diet, muscle function and mRNA levels of mitochondrial UPRmt pathway proteins decreased significantly. Four weeks of aerobic training increased grip strength, maximum speed, along with improving mRNA levels of Lon protease homolog, mitochondrial (LONP1), HSP60 ClpP, which shows that aerobic exercise can reduce the negative effects of obesity on skeletal muscle by the function increasing of the UPRmt pathway(24). The UPRmt response is one of the quality control mechanisms of MQC, which uses Activating Transcription Factor 5 (ATF5) to induce the expression of protective enzymes to maintain mitochondrial function. An increase in ATF5 expression after a bout of aerobic activity has been reported. This shows that the transcription factor ATF5 plays a crucial role in maintaining mitochondrial homeostasis and the proper response of muscle to physical activity to optimize mitochondrial quality control (25). In the hypothalamus, aerobic exercise increased the UPRmt markers of the hypothalamus. This was with maximum mitochondrial associated respiratory capacity in the brain, which confirms the role of aerobic exercise in the activation of the UPRmt pathway (26). Li et al. (2023)

reviewed the role of regular physical activities in mitochondrial preserving proteostasis in Parkinson's disease. The results of this study showed that regular physical activities maintain mitochondrial proteostasis by activating UPRmt, mitophagy. It regulates mitochondrial protein levels by regulating mitochondrial protein reducing oxidative transport, stress and improving bodv metabolism. energy Mitochondrial proteostasis regulation prevents and improves Parkinson's disease (27).

The effect of aerobic exercise on **Mitochondrial dynamics (fission and** fusion)

Mitochondrial dynamics includes the process of changing the morphology, quantity and position of mitochondria in eukaryotic cells, which is of great importance for the proper functioning of including the cell. energy production, differentiation, cell cycle, aging and apoptosis (28). Mitochondrial dynamics are vital in regulating many vital cell processes and maintaining mitochondrial homeostasis in response to stress (29,30). Mitochondrial dynamics consists of two opposite processes called fission and fusion. While fission separates mitochondria into globular organelles (31), fusion allows mitochondrial components to regenerate into an expanded mitochondrial network (32), facilitating mitochondrial metabolic remodeling and quality control. Disturbance in mitochondrial dynamics is one of the key mechanisms in the pathogenesis of many diseases such as diabetes, heart failure, Parkinson's disease, Huntington's disease, and cardiac hypertrophy, heart failure, myocardial infarction and ischemia-amyotrophic lateral sclerosis, in which mitochondrial dysfunction plays a role in all these diseases. Is important (33,34).

Physiological conditions cause mitochondria to adapt to cellular energy needs. These changes can occur through continuous cycles of mitochondrial fusion and fission, which allows for an adequate distribution of mitochondria within cells. Mitochondrial fission produces small mitochondria, while large interconnected mitochondria networks are produced through fusion (35). Mitochondrial fusion and fission are necessary to maintain critical cellular functions including mitochondrial respiratory activity, mitochondrial DNA (mtDNA) distribution. apoptosis and cell survival (36). It has been reported that twelve weeks of high-intensity aerobic interval training increased the volume and number of skeletal muscle mitochondria along with an increase in mitochondrial respiration and insulin sensitivity. OPA1, a regulator of fusion, was significantly increased following HIIT while FIS1, a regulator of fission, was significantly decreased following HIIT. The OPA1/FIS1 ratio was also significantly increased, indicating a balance between fission and fusion. This was positively correlated with improved respiration, insulin sensitivity, etc. Changes indicate increased fusion following HIIT with increased mitochondrial respiration, insulin sensitivity, and Vo2peak. This supports the idea that increased mitochondrial fusion is associated with significant health benefits of HIIT (37). In another study, twelve weeks of aerobic exercise improved insulin sensitivity, aerobic capacity, and fatty acid oxidation. Aerobic exercise decreased FIS1 and Parkin (regulator of fission) in skeletal muscle, while significantly affecting the expression of MFN1, MFN2, OPA1, and OMA1. (Fusion regulator) did not have. Aerobic exercise improves the ratio of fusion to fission proteins. These findings indicate that aerobic training changes the expression of mitochondrial fission and fusion proteins, and these changes may enhance insulin sensitivity and reduce metabolic disorders (38).

Mitochondrial dynamics are improved by aerobic exercise. Exercises that are very intense can reduce intrinsic mitochondrial function. It has been shown that aerobic exercise is beneficial depending on the intensity and duration of the program (40). In their review, Heo et al. (2017) showed that obesity disrupts mitochondrial dynamics and leads to an imbalance between fusion and fission by favoring fission or decreasing fusion proteins. Mitochondrial dysfunction and oxidative stress induce apoptosis, and mitochondrial apoptosis is induced by obesity in skeletal muscle. It is well known that regular physical activity is the most effective intervention to protect against obesity. reduce Regular physical activity can dysfunction mitochondrial and improve mitochondrial dynamics by reducing oxidative stress (41). Campos et al. (2023) showed that mitochondrial connectivity AND cycle dynamics are necessary to maintain physical fitness during aging. Regular physical activity can moderate aging's negative effects on skeletal muscle by regulating mitochondrial proteins (29).

Effect of aerobic exercise on Mitochondrial mitophagy

Mitophagy is selective autophagy of damaged mitochondria (42). Mitophagy is a multi-step process that is very complexly regulated and destruction causes the selective of damaged/dysfunctional mitochondria bv autophagy and has common aspects with other types of selective autophagy. From a morphological point of view, mitophagy is the placement of mitochondria inside an autophagic vacuole called a mitophagosome (43). In general, mitophagy is a fundamental biological mechanism in all cells or tissues and is regulated by cellular energy needs.

Mitophagy amounts are different in all tissues. In tissues such as the nervous system, kidney, skeletal muscle, heart and liver, it is more than the spleen and thymus (44).

Biological evidence shows that damaged mitochondria are not only unable to produce ATP and other biosynthetic products, but also produce higher levels of reactive oxygen species. This can disturb the intracellular balance. If reactive oxygen species cannot be cleared in time and accumulate in cells, it leads to pathological apoptosis (45).

Mitophagy keeps mitochondria in optimal condition by removing dysfunctional mitochondria. Mitochondrial homeostasis is maintained by the balance between elimination and bioproduction, which can be disrupted by uncontrolled mitophagy (46). Disruption in mitophagy can be associated with diseases such as type 2 diabetes, non-alcoholic fatty liver disease. cardiovascular diseases. neurodegenerative diseases and old age (47). PTEN-induced putative kinase 1 (PINK1) and Parkin (also known as PARK2) are two key proteins that play a crucial role in mitochondrial quality control (48). PINK1 is а serine/threonine kinase located in mitochondria. cells It protects from mitochondrial dysfunction and stress. When mitochondria are damaged, PINK1 accumulates in the outer membrane of dysfunctional mitochondria. PINK1 acts as a signal for the selective destruction of damaged mitochondria through mitophagy. It attracts Parkin to the damaged mitochondria and removes these organelles (49). Parkin is an E3 ubiquitin ligase that regulates protein degradation through the ubiquitin-proteasome system. It also controls mitochondria and mitophagy (50). When PINK1 accumulates in damaged mitochondria, it phosphorylates Parkin and activates it.

Upon activation, Parkin ubiquitinates various proteins on the mitochondrial outer membrane. This marks them for degradation and facilitates the recruitment of the autophagic machinery to engulf and destroy damaged mitochondria (51). Like UPRmt and mitochondrial dynamics, aerobic exercise positively affects mitophagy in aging, obesity and various diseases. It has been reported that aerobic exercise increases the proteins of the mitophase pathway, especially Parkin, along with the improvement of mitochondrial function in the myocardium (52). On the other hand, aerobic exercise reduced age-related muscle mass destruction in rodents by activating the PINK/Parkin signaling pathway and regulating mitophagy (53). These findings have also been reported in old rats' myocardial tissue after aerobic exercise (54). An increase in the expression of PINK/Parkin protein with an increase in mitophagy, improving mitochondrial function, increasing the volume and number of skeletal muscle mitochondria and aerobic performance after aerobic exercise has also been reported (55). High-intensity interval training significantly increased mitochondrial biogenesis and mitophagy in parallel with reducing inflammation and oxidative stress in skeletal muscle. These changes were associated with an increase in fatigue time (56). These findings have been confirmed in the skeletal muscle of obese elderly people after intermittent exercise with high intensity, which indicates the regulatory effect of aerobic exercise on mitophagy (57).

Conclusion

The studies review of showed that mitochondrial health is directly related to cell function. In addition, health and mitochondrial disorders are the pathogenesis of many diseases such as metabolic diseases, neurodegenerative disorders, and cancer. Aerobic exercise can develop mitochondrial quality control and improve health and physical performance by affecting the three mechanisms of mitochondrial unfolded protein response (UPRmt), mitochondrial dynamics (fission and fusion) and mitochondrial mitophagy. In fact, aerobic exercise enhances health by increasing the levels of mRNA and proteins responsible for the signaling pathway (UPRmt), mitochondrial dynamics (fission and fusion) and mitochondrial mitophagy.

Acknowledgements

Hereby, from all the patients and people participating in the present research and their loved ones We are grateful to those who have helped us in this research.

Funding

This study did not have any funds.

Compliance with ethical standards

Conflict of interest None declared.

Ethical approval the research was conducted with regard to the ethical principles.

Informed consent Informed consent was obtained from all participants.

Author contributions

Conceptualization: F,A. M,A,A. SH,R,M. L,M ; Methodology: F,A. M,A,A. SH,R,M. L,M ; Software: F,A. M,A,A. SH,R,M. L,M ; Validation: F,A. M,A,A. SH,R,M. L,M ;Formal analysis: F,A. M,A,A. SH,R,M. L,M ;Investigation: F,A. M,A,A. SH,R,M. L,M ; Resources: F,A. M,A,A. SH,R,M. L,M ;Data curation: F,A. M,A,A. SH,R,M. L,M ; Writing - original draft: M.H, M.G, R.Z, P.E; Writing review & editing: M.H, M.G, R.Z, P.E; Visualization: F,A. M,A,A. SH,R,M. L,M; Supervision: F,A. M,A,A. SH,R,M. L,M ;Project administration: F,A. M,A,A. SH,R,M. L,M; Funding acquisition: F,A. M,A,A. SH,R,M. L,M.

References

1. Almutairi AH, Almutairi NS, Mousa N, Elsayed A, El-Sehrawy A, Elmetwalli A. Aerobic exercise as a non-pharmacological intervention for improving metabolic and hemodynamic profiles in type 2 diabetes. Ir J Med Sci. 2024 Dec;193(6):2781-2790. doi: 10.1007/s11845-024-03783-6. Epub 2024 Aug 19. PMID: 39158674.

2. Riyahi Malayeri, S., Mirakhorli, M. The Effect of 8 Weeks of Moderate Intensity Interval Training on Omentin Levels and Insulin Resistance Index in Obese Adolescent Girls. Sport Physiology & Management Investigations, 2018; 10(2): 59-68.

3. Choi JW, Jo SW, Kim DE, Paik IY, Balakrishnan R. Aerobic exercise attenuates LPS-induced cognitive dysfunction by reducing oxidative stress, glial activation, and neuroinflammation. Redox Biol. 2024 May;71:103101.

4. Farazandeh Nia, D., Hosseini, M., Riyahi Malayeri, S., Daneshjoo, A. Effect of Eight Weeks of Swimming Training with Garlic Intake on Serum Levels of IL-10 and TNF- α in Obese Male Rats. Jundishapur Scientific Medical Journal, 2018; 16(6): 665-671. doi: 10.22118/jsmj.2018.57830.

5. Bao F, Zhao X, You J, Liu Y, Xu Z, Wu Y, Wu Y, Xu Z, Yu L, Li J, Wei Y. Aerobic exercise alleviates skeletal muscle aging in male rats by inhibiting apoptosis via regulation of the Trx system. Exp Gerontol. 2024 Sep;194:112523. doi: 10.1016/j.exger.2024.112523. Epub 2024 Jul 18. Erratum in: Exp Gerontol. 2024 Oct 1;195:112541. doi: 10.1016/j.exger.2024.112541. PMID: 39025384.

6. Chen Q, Zhao X, Xu Z, Liu Y. Endoplasmic reticulum stress mechanisms and exercise intervention in type 2 diabetes mellitus. Biomed Pharmacother. 2024 Aug;177:117122. doi: 10.1016/j.biopha.2024.117122. Epub 2024 Jul 10. PMID: 38991302.

7. Zhang F, Lin JJ, Tian HN, Wang J. Effect of exercise on improving myocardial mitochondrial function in decreasing diabetic cardiomyopathy. Exp Physiol. 2024 Feb;109(2):190-201. doi: 10.1113/EP091309. Epub 2023 Oct 16. PMID: 37845840; PMCID: PMC10988701.

8. Javadov S, Kozlov AV, Camara AKS. Mitochondria in Health and Diseases. Cells. 2020 May 9;9(5):1177. doi: 10.3390/cells9051177. PMID: 32397376; PMCID: PMC7290976. 9. Mookerjee SA, Goncalves RLS, Gerencser AA, Nicholls DG, Brand MD. The contributions of respiration and glycolysis to extracellular acid production. Biochim Biophys Acta. 2015 Feb;1847(2):171-181. doi: 10.1016/j.bbabio.2014.10.005. Epub 2014 Oct 27. PMID: 25449966.

10. Picard M, Wallace DC, Burelle Y. The rise of mitochondria in medicine. Mitochondrion. 2016 Sep;30:105-16. doi: 10.1016/j.mito.2016.07.003. Epub 2016 Jul 14. PMID: 27423788; PMCID: PMC5023480.. Epub 2015 Aug 20. PMID: 26298752.

11. Hosseini M, Ghasem Zadeh Khorasani N, Divkan B, Riyahi Malayeri S. Interactive Effect of High Intensity Interval Training with Vitamin E Consumption on the Serum Levels of Hsp70 and SOD in Male Wistar Rats. Iranian J Nutr Sci Food Technol 2019; 13 (4) :21-28

URL: http://nsft.sbmu.ac.ir/article-1-2689-en.html

12. Lightowlers RN, Taylor RW, Turnbull DM. Mutations causing mitochondrial disease: What is new and what challenges remain? Science. 2015 Sep 25;349(6255):1494-9. doi: 10.1126/science.aac7516. Epub 2015 Sep 24. PMID: 26404827.

13. Tanaka T, Nishimura A, Nishiyama K, Goto T, Numaga-Tomita T, Nishida M. Mitochondrial dynamics in exercise physiology. Pflugers Arch. 2020 Feb;472(2):137-153. doi: 10.1007/s00424-019-02258-3. Epub 2019 Feb 1. PMID: 30707289.

14. Zeng Z, Liang J, Wu L, Zhang H, Lv J, Chen N. Exercise-Induced Autophagy Suppresses Sarcopenia Through Akt/mTOR and Akt/FoxO3a Signal Pathways and AMPK-Mediated Mitochondrial Quality Control. Front Physiol. 2020 Nov 2;11:583478. doi: 10.3389/fphys.2020.583478. PMID: 33224037; PMCID: PMC7667253.

15. Gao F, Zhang J. Mitochondrial quality control and neurodegenerative diseases. Neuronal Signal. 2018 Dec 3;2(4):NS20180062. doi: 10.1042/NS20180062. PMID: 32714594; PMCID: PMC7373240.

16. Hu D, Liu Z, Qi X. Mitochondrial Quality Control Strategies: Potential Therapeutic Targets for Neurodegenerative Diseases? Front Neurosci. 2021 Nov 12;15:746873. doi: 10.3389/fnins.2021.746873. PMID: 34867159; PMCID: PMC8633545. 17. Inigo JR, Chandra D. The mitochondrial unfolded protein response (UPR^{mt}): shielding against toxicity to mitochondria in cancer. J Hematol Oncol. 2022 Jul 21;15(1):98. doi: 10.1186/s13045-022-01317-0. PMID: 35864539; PMCID: PMC9306209.

 Kenny TC, Craig AJ, Villanueva A, Germain D. Mitohormesis Primes Tumor Invasion and Metastasis. Cell Rep. 2019 May 21;27(8):2292-2303.e6. doi: 10.1016/j.celrep.2019.04.095. PMID: 31116976; PMCID: PMC6579120.

19. Cole A, Wang Z, Coyaud E, Voisin V, Gronda M, Jitkova Y, Mattson R, Hurren R, Babovic S, Maclean N, Restall I, Wang X, Jeyaraju DV, Sukhai MA, Prabha S, Bashir S, Ramakrishnan A, Leung E, Qia YH, Zhang N, Combes KR, Ketela T, Lin F, Houry WA, Aman A, Al-Awar R, Zheng W, Wienholds E, Xu CJ, Dick J, Wang JC, Moffat J, Minden MD, Eaves CJ, Bader GD, Hao Z, Kornblau SM, Raught B, Schimmer AD. Inhibition of the Mitochondrial Protease ClpP as a Therapeutic Strategy for Human Acute Myeloid Leukemia. Cancer Cell. 2015 Jun 8;27(6):864-76. doi: 10.1016/j.ccell.2015.05.004. PMID: 26058080; PMCID: PMC4461837.

20. Hood DA, Memme JM, Oliveira AN, Triolo M. Maintenance of Skeletal Muscle Mitochondria in Health, Exercise, and Aging. Annu Rev Physiol. 2019 Feb 10;81:19-41. doi: 10.1146/annurev-physiol-020518-114310. Epub 2018 Sep 14. PMID: 30216742.

21. Wang Z, Bo H, Song Y, Li C, Zhang Y. Mitochondrial ROS Produced by Skeletal Muscle Mitochondria Promote the Decisive Signal for UPRmt Activation. Biomed Res Int. 2022 Feb 21;2022:7436577. doi: 10.1155/2022/7436577. Retraction in: Biomed Res Int. 2023 Dec 29;2023:9898164. doi: 10.1155/2023/9898164. PMID: 35237690; PMCID: PMC8885241.

22. Cordeiro AV, Brícola RS, Braga RR, Lenhare L, Silva VRR, Anaruma CP, Katashima CK, Crisol BM, Simabuco FM, Silva ASR, Cintra DE, Moura LP, Pauli JR, Ropelle ER. Aerobic Exercise Training Induces the Mitonuclear Imbalance and UPRmt in the Skeletal Muscle of Aged Mice. J Gerontol A Biol Sci Med Sci. 2020 Nov 13;75(12):2258-2261. doi: 10.1093/gerona/glaa059. PMID: 32173728.

23. Cordeiro AV, Peruca GF, Braga RR, Brícola RS, Lenhare L, Silva VRR, Anaruma CP, Katashima CK, Crisol BM, Barbosa LT, Simabuco FM, da Silva ASR, Cintra DE, de Moura LP, Pauli JR, Ropelle ER. High-intensity exercise training induces mitonuclear imbalance and activates the mitochondrial unfolded protein response in the skeletal muscle of aged mice. Geroscience. 2021 Jun;43(3):1513-1518. doi: 10.1007/s11357-020-00246-5. Epub 2020 Jul 31. PMID: 32737758; PMCID: PMC8190321.

24. Apablaza P, Bórquez JC, Mendoza R, Silva M, Tapia G, Espinosa A, Troncoso R, Videla LA, Juretić N, Del Campo A. Exercise Induces an Augmented Skeletal Muscle Mitochondrial Unfolded Protein Response in a Mouse Model of Obesity Produced by a High-Fat Diet. Int J Mol Sci. 2023 Mar 16;24(6):5654. doi: 10.3390/ijms24065654. PMID: 36982728; PMCID: PMC10051316.

25. Slavin MB, Kumari R, Hood DA. ATF5 is a regulator of exercise-induced mitochondrial quality control in skeletal muscle. Mol Metab. 2022 Dec;66:101623. doi: 10.1016/j.molmet.2022.101623. Epub 2022 Nov 1. PMID: 36332794; PMCID: PMC9661517.

26 . Braga RR, Crisol BM, Brícola RS, et al. Exercise alters the mitochondrial proteostasis and induces the mitonuclear imbalance and UPR^{mt} in the hypothalamus of mice. Scientific Reports. 2021 Feb;11(1):3813. DOI: 10.1038/s41598-021-82352-8. PMID: 33589652; PMCID: PMC7884690.

27. Li J, Xu Y, Liu T, Xu Y, Zhao X, Wei J. The Role of Exercise in Maintaining Mitochondrial Proteostasis in Parkinson's Disease. Int J Mol Sci. 2023 Apr 28;24(9):7994. doi: 10.3390/ijms24097994. PMID: 37175699; PMCID: PMC10179072.

28. Chen W, Zhao H, Li Y. Mitochondrial dynamics in health and disease: mechanisms and potential targets. Signal Transduct Target Ther. 2023 Sep 6;8(1):333. doi: 10.1038/s41392-023-01547-9. PMID: 37669960; PMCID: PMC10480456.

29. Campos JC, Marchesi Bozi LH, Krum B, Grassmann Bechara LR, Ferreira ND, Arini GS, Albuquerque RP, Traa A, Ogawa T, van der Bliek AM, Beheshti A, Chouchani ET, Van Raamsdonk JM, Blackwell TK, Ferreira JCB. Exercise preserves physical fitness during aging through AMPK and mitochondrial dynamics. Proc Natl Acad Sci U S A. 2023 Jan 10;120(2):e2204750120

30. Meyer JN, Leuthner TC, Luz AL. Mitochondrial fusion, fission, and mitochondrial toxicity. Toxicology. 2017 Nov 1;391:42-53. doi: 10.1016/j.tox.2017.07.019. Epub 2017 Aug 5. PMID: 28789970; PMCID: PMC5681418.

31. Twig G, Elorza A, Molina AJ, Mohamed H, Wikstrom JD, Walzer G, Stiles L, Haigh SE, Katz S, Las G, Alroy J, Wu M, Py BF, Yuan J, Deeney JT, Corkey BE, Shirihai OS. Fission and selective fusion govern mitochondrial segregation and elimination by autophagy. EMBO J. 2008 Jan 23;27(2):433-46. doi: 10.1038/sj.emboj.7601963. Epub 2008 Jan 17. PMID: 18200046; PMCID: PMC2234339.

32. Chen H, Chomyn A, Chan DC. Disruption of fusion results in mitochondrial heterogeneity and dysfunction. J Biol Chem.
2005 Jul 15;280(28):26185-92. doi: 10.1074/jbc.M503062200. Epub 2005 May 17. PMID: 15899901.

33. Suárez-Rivero JM, Villanueva-Paz M, de la Cruz-Ojeda P, de la Mata M, Cotán D, Oropesa-Ávila M, de Lavera I, Álvarez-Córdoba M, Luzón-Hidalgo R, Sánchez-Alcázar JA. Mitochondrial Dynamics in Mitochondrial Diseases. Diseases. 2016 Dec 23;5(1):1. doi: 10.3390/diseases5010001. PMID: 28933354; PMCID: PMC5456341.

34. Yapa NMB, Lisnyak V, Reljic B, Ryan MT. Mitochondrial dynamics in health and disease. FEBS Lett. 2021 Apr;595(8):1184-1204. doi: 10.1002/1873-3468.14077. Epub 2021 Apr 5. PMID: 33742459.

35. Youle RJ, van der Bliek AM. Mitochondrial fission, fusion, and stress. Science. 2012 Aug 31;337(6098):1062-5. doi: 10.1126/science.1219855. PMID: 22936770; PMCID: PMC4762028.

36. Westermann B. Mitochondrial fusion and fission in cell life and death. Nat Rev Mol Cell Biol. 2010 Dec;11(12):872-84. doi: 10.1038/nrm3013. PMID: 21102612.

37. Ruegsegger GN, Pataky MW, Simha S, Robinson MM, Klaus KA, Nair KS. High-intensity aerobic, but not resistance or combined, exercise training improves both cardiometabolic health and skeletal muscle mitochondrial dynamics. J Appl Physiol (1985). 2023 Oct 1;135(4):763-774. doi: 10.1152/japplphysiol.00405.2023. Epub 2023 Aug 24. PMID: 37616334; PMCID: PMC10642518.

38. Axelrod CL, Fealy CE, Mulya A, Kirwan JP. Exercise training remodels human skeletal muscle mitochondrial fission and fusion machinery towards a pro-elongation phenotype. Acta Physiol (Oxf). 2019 Apr;225(4):e13216. doi: 10.1111/apha.13216. Epub 2018 Dec 1. PMID: 30408342; PMCID: PMC6416060.

39. Moore TM, Zhou Z, Cohn W, et al. The impact of exercise on mitochondrial dynamics and the role of Drp1 in exercise performance and training adaptations in skeletal muscle. Molecular Metabolism. 2019 Mar;21:51-67. DOI: 10.1016/j.molmet.2018.11.012. PMID: 30591411; PMCID: PMC6407367.

40. Flockhart M, Nilsson LC, Tais S, Ekblom B, Apró W, Larsen FJ. Excessive exercise training causes mitochondrial functional impairment and decreases glucose tolerance in healthy volunteers. Cell Metab. 2021 May 4;33(5):957-970.e6. doi: 10.1016/j.cmet.2021.02.017. Epub 2021 Mar 18. PMID: 33740420.

41. Heo JW, No MH, Park DH, Kang JH, Seo DY, Han J, Neufer PD, Kwak HB. Effects of exercise on obesity-induced mitochondrial dysfunction in skeletal muscle. Korean J Physiol Pharmacol. 2017 Nov;21(6):567-577. doi: 10.4196/kjpp.2017.21.6.567. Epub 2017 Oct 30. PMID: 29200899; PMCID: PMC5709473.

42. Ding WX, Yin XM. Mitophagy: mechanisms, pathophysiological roles, and analysis. Biol Chem. 2012 Jul;393(7):547-64. doi: 10.1515/hsz-2012-0119. PMID: 22944659; PMCID: PMC3630798.

43. Zachari M, Ktistakis NT. Mammalian Mitophagosome Formation: A Focus on the Early Signals and Steps. Front Cell Dev Biol. 2020 Mar 18;8:171. doi: 10.3389/fcell.2020.00171. PMID: 32258042; PMCID: PMC7093328.

44. Cummins N, Götz J. Shedding light on mitophagy in neurons: what is the evidence for PINK1/Parkin mitophagy in vivo? Cell Mol Life Sci. 2018 Apr;75(7):1151-1162. doi: 10.1007/s00018-017-2692-9. Epub 2017 Oct 30. PMID: 29085955; PMCID: PMC11105538.

45. Cadenas E, Davies KJ. Mitochondrial free radical generation, oxidative stress, and aging. Free Radic Biol Med. 2000 Aug;29(3-4):222-30. doi: 10.1016/s0891-5849(00)00317-8. PMID: 11035250.

46. Ashrafi, G., Schwarz, T. The pathways of mitophagy for quality control and clearance of mitochondria. *Cell Death Differ* **20**, 31–42 (2013). https://doi.org/10.1038/cdd.2012.81

47. Picca A, Faitg J, Auwerx J, Ferrucci L, D'Amico D. Mitophagy in human health, ageing and disease. Nat Metab. 2023 Dec;5(12):2047-2061. doi: 10.1038/s42255-023-00930-8. Epub 2023 Nov 30. PMID: 38036770.

48. Pickrell AM, Youle RJ. The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. Neuron. 2015 Jan 21;85(2):257-73. doi: 10.1016/j.neuron.2014.12.007. PMID: 25611507; PMCID: PMC4764997.

49. Quinn PMJ, Moreira PI, Ambrósio AF, Alves CH. PINK1/PARKIN signalling in neurodegeneration and neuroinflammation. Acta Neuropathol Commun. 2020 Nov 9;8(1):189. doi: 10.1186/s40478-020-01062-w. PMID: 33168089; PMCID: PMC7654589.

50. Ge, P., Dawson, V.L. & Dawson, T.M. PINK1 and Parkin mitochondrial quality control: a source of regional vulnerability in Parkinson's disease. *Mol Neurodegeneration* **15**, 20 (2020). https://doi.org/10.1186/s13024-020-00367-7

Research article

51. Jin SM, Youle RJ. PINK1- and Parkin-mediated mitophagy at a glance. J Cell Sci. 2012 Feb 15;125(Pt 4):795-9. doi: 10.1242/jcs.093849. PMID: 22448035; PMCID: PMC3656616.

52. Nijholt, K. T., Sánchez-Aguilera, P. I., Mahmoud, B., Gerding, A., Wolters, J. C., Wolters, A. H. G., Giepmans, B. N. G., Silljé, H. H. W., de Boer, R. A., Bakker, B. M., & Westenbrink, B. D. (2023). A Kinase Interacting Protein 1 regulates mitochondrial protein levels in energy metabolism and promotes mitochondrial turnover after exercise. *Scientific Reports*, *13*, Article 18822. https://doi.org/10.1038/s41598-023-45961-z

53. Chen YL, Ma YC, Tang J, Zhang D, Zhao Q, Liu JJ, Tang HS, Zhang JY, He GH, Zhong CH, Wu YT, Wen HR, Ma LQ, Zou CG. Physical exercise attenuates age-related muscle atrophy and exhibits anti-ageing effects via the adiponectin receptor 1 signalling. J Cachexia Sarcopenia Muscle. 2023 Aug;14(4):1789-1801.

54. No MH, Heo JW, Yoo SZ, Kim CJ, Park DH, Kang JH, Seo DY, Han J, Kwak HB. Effects of aging and exercise training on mitochondrial function and apoptosis in the rat heart. Pflugers Arch. 2020 Feb;472(2):179-193. doi: 10.1007/s00424-020-02357-6. Epub 2020 Feb 11. PMID: 32048000.

55. Ma C, Zhao Y, Ding X, Gao B. The role of Sirt3 in the changes of skeletal muscle mitophagy induced by hypoxic training. Gen Physiol Biophys. 2022 Sep;41(5):447-455. doi: 10.4149/gpb_2022023. PMID: 36222342.

56. Yamauchi N, Tamai K, Kimura I, Naito A, Tokuda N, Ashida Y, Motohashi N, Aoki Y, Yamada T. High-intensity interval training in the form of isometric contraction improves fatigue resistance in dystrophin-deficient muscle. J Physiol. 2023 Jul;601(14):2917-2933. doi: 10.1113/JP284532. Epub 2023 May 22. PMID: 37184335.